

TRICHOHECENE RESISTANCE GENE

This application is a continuation of copending U.S. Application No. 10/074,279, filed February 12, 2002, which claims benefit of U.S. Application No. 09/538,414, filed March 29, 2000, now U.S. Patent No. 6,346,655, which claims the benefit of U.S. Provisional Application No. 60/304,177, filed March 31, 1999, and U.S. Provisional Application No. 60/287,549, filed February 11, 2000, all of which are incorporated herein by reference in their entirety.

A) Subject Matter of the Invention

The present invention relates to transgenic hosts particularly transgenic plants, plant tissues, seeds and cells that are trichothecene resistant and methods of making and using the same. The present invention further relates to methods of preventing and/or reducing fungal growth on a plant, plant tissue, seed or plant cell. The present invention further relates to preventing and/or reducing mycotoxin contamination of a plant, plant tissue or seed. The present invention further relates to using trichothecenes as selective agents in transformation protocols.

B) Background

Numerous fungi are serious pests of economically important agricultural crops. Further, crop contamination by fungal toxins is a major problem for agriculture throughout the world. Mycotoxins are toxic fungal metabolites, often found in agricultural products that are characterized by their ability to cause health problems for vertebrates. Trichothecenes are sesquiterpene epoxide mycotoxins produced by species of *Fusarium*, *Trichothecium*, and *Myrothecium* that act as potent inhibitors of eukaryotic protein synthesis. *Fusarium* species that produce such trichothecenes include *F. acuminatum*, *F. crookwellense*, *F. culmorum*, *F. equiseti*, *F. graminearum* (*Gibberella zeae*), *F. lateritium*, *F. poae*, *F. sambucinum* (*G. pulicaris*), and *F. sporotrichioides* (Marasas, W.F.O., Nelson, P.E., and Toussoun, T.A. 1984).

As previously described (A. E.Desjardins and T. M Hohn, Mycotoxins in plant pathogenesis.Mol.Plant-Microbe Interact. 10 (2):147-152, 1997), both acute and chronic mycotoxicoses in farm animals and in humans have been associated with consumption of wheat, rye, barley, oats, rice and maize contaminated with *Fusarium* species that produce

trichothecene mycotoxins. Experiments with chemically pure trichothecenes at low dosage levels have reproduced many of the features observed in moldy-grain toxicoses in animals, including anemia and immunosuppression, hemorrhage, emesis and feed refusal. Historical and epidemiological data from human populations indicate an association between certain disease epidemics and consumption of grain infected with *Fusarium* species that produce trichothecenes. In particular, outbreaks of a fatal disease known as alimentary toxic aleukia, which has occurred in Russia since the nineteenth century, have been associated with consumption of over-wintered grains contaminated with *Fusarium* species that produce the trichothecene T-2 toxin. In Japan, outbreaks of a similar disease called akakabi-byo or red mold disease have been associated with grain infected with *Fusarium* species that produce the trichothecene, deoxynivalenol (hereinafter "DON"). Trichothecenes were detected in the toxic grain samples responsible for recent human disease outbreaks in India and Japan. There exists, therefore, a need for agricultural methods for preventing and, crops having reduced levels of, mycotoxin contamination.

Further, trichothecene-producing *Fusarium* species are destructive pathogens and attack a wide range of plant species. The acute phytotoxicity of trichothecenes and their occurrence in plant tissues also suggest that these mycotoxins play a role in the pathogenesis of *Fusarium* on plants. This implies that mycotoxins play a role in disease and, therefore, reducing their toxicity to the plant may also prevent or reduce disease in the plant. Further, reduction in disease levels may have the additional benefit of reducing mycotoxin contamination on the plant and particularly in grain where the plant is a cereal plant.

Various methods of controlling diseases in plants, such as corn ear rot, stock rot or wheat head blight, have been used with varying degrees of success. One method of controlling plant disease has been to apply an antimicrobial chemical to crops. This method has numerous, art-recognized problems. Alternatively, a more recent method involves the use of biological control organisms ("biocontrol") which are natural competitors or inhibitors of the pest organism. However, it is difficult to apply biocontrol to large areas, and even more difficult to cause those living organisms to remain in the treated area for an extended period of time. More recently, techniques in recombinant DNA have provided the opportunity to insert into plant cells cloned genes, which express antimicrobial compounds. However, this technology has given rise to concerns about eventual microbial resistance to well-known, naturally occurring antimicrobials. Thus, a continuing need exists to identify naturally

occurring antimicrobial agents, such as proteins, which can be formed by plant cells directly by translation of a single gene.

A trichothecene 3-O-acetyltransferase that catalyzes the acetylation of a number of different *Fusarium* trichothecenes including DON at the C3 hydroxyl group has been identified in *Fusarium sporotrichioides*. (S. P. McCormick, N. J. Alexander, S. C. Trapp, and T. M. Hohn. Disruption of TRI101, the gene encoding trichothecene 3-O-acetyltransferase, from *Fusarium sporotrichioides*. *Applied Environ. Microbiol.* 65 (12):5252-5256, 1999.) Acetylation of trichothecenes at the C3-OH significantly reduces their toxicity in vertebrates and plants and results in the reaction product 3-acetyldeoxynivalenol (hereinafter "3ADON") See, Kimura et al. below.

The sequence of structural genes encoding trichothecene 3-O-acetyl transferases from *Fusarium graminearum*, *Fusarium sporotrichioides* as well as sequences of other orthologs has been published. See, e.g. Kimura et al., *Biosci. Biotechnol. Biochem.*, 62 (5) 1033-1036 (1998), and Kimura et al., *FEBS Letters*, 435, 163-168 (1998). Further, it has been speculated that the gene from *Fusarium sporotrichioides* encoding a trichothecene 3-O-acetyl transferase may be useful in developing plant varieties with increased resistance to *Fusarium*. See., e.g. Hohn, T.M. et al. *Molecular Genetics of Host-Specific Toxins in Plant Disease*, 17-24 (1998), and Kimura et al. *J.Biological Chemistry*, 273(3) 1654-1661 (1998).

Prior to the present invention, however, many uncertainties rendered it far from obvious whether expressing trichothecene 3-O-acetyl transferases in a plant would actually lead to trichothecene resistant plants. For example, the reaction catalyzed by the *Fusarium sporotrichioides* trichothecene 3-O-acetyl transferase is reversible and might, therefore have failed to protect plant cells from trichothecenes such as DON. It was also uncertain whether there might be esterases in plant cells that would compete with the 3-O-acetyl transferase activities to generate toxic DON from 3ADON. It was also uncertain how the metabolism of the reaction product 3ADON might affect the plant, e.g. whether introduction of the trichothecene 3-O- acetyltransferase would alter plant growth and development in ways that would negate any positive contribution of the acetyltransferase by for example, interfering with the plant's natural disease resistance mechanisms. It was also uncertain whether 3ADON could be metabolized by the plant to form a novel secondary metabolite with toxic effects. It was also uncertain, even if DON produced by an invading fungus was efficiently converted to

3ADON, whether this conversion would impart enhanced pathogen resistance upon the plant. The above are but a few of the uncertainties in the art before the time of the present invention.

C) Definitions

Expression refers to the transcription and/or translation of an endogenous gene or a transgene in plants. In the case of antisense constructs, for example, expression may refer to the transcription of the antisense DNA only.

Operably linked/associated when referring to a regulatory DNA sequence being "operably linked to" or "associated with" a DNA sequence that codes for an RNA or a protein refers to the two sequences being situated such that the regulatory DNA sequence affects expression of the coding DNA sequence.

The term "heterologous polynucleotide" or "heterologous DNA" as used herein each refers to a nucleic acid molecule not naturally associated with a host cell into which it is introduced, including genetic constructs, non-naturally occurring multiple copies of a naturally occurring nucleic acid molecule; and an otherwise homologous nucleic acid molecule operatively linked to a non-native nucleic acid molecule. Thus, a heterologous gene in a host cell includes a gene that is endogenous to the particular host cell but has been modified through, for example, the use of DNA shuffling. Thus, the term encompasses a DNA segment that is foreign or heterologous to the cell, or homologous to the cell but in a position within the host cell nucleic acid in which the element is not ordinarily found.

The terms "nucleic acid" or "polynucleotide" refers to deoxyribonucleotides or ribonucleotides and polymers thereof in either single- or double-stranded form. Unless specifically limited, the term encompasses nucleic acids containing known analogues of natural nucleotides which have similar binding properties as the reference nucleic acid and are metabolized in a manner similar to naturally occurring nucleotides. Unless otherwise indicated, a particular nucleic acid sequence also implicitly encompasses conservatively modified variants thereof (*e.g.* degenerate codon substitutions) and complementary sequences and as well as the sequence explicitly indicated. Specifically, degenerate codon substitutions may be achieved by generating sequences in which the third position of one or more selected (or all) codons is substituted with mixed-base and/or deoxyinosine residues (Batzer *et al.*, *Nucleic Acid Res.* 19: 5081 (1991); Ohtsuka *et al.*, *J. Biol. Chem.* 260: 2605-2608 (1985); Rossolini *et al.*, *Mol. Cell. Probes* 8: 91-98 (1994)). The terms "nucleic acid" or "nucleic acid

sequence" or "polynucleotide" may also be used interchangeably with gene, cDNA, and mRNA encoded by a gene.

In its broadest sense, the term "substantially similar", when used herein with respect to a nucleic acid molecule, means a nucleic acid molecule corresponding to a reference nucleotide sequence, wherein the corresponding nucleic acid molecule encodes a polypeptide having substantially the same structure and function as the polypeptide encoded by the reference nucleotide sequence, e.g. where only changes in amino acids not affecting the polypeptide function occur. Desirably the substantially similar nucleic acid molecule encodes the polypeptide encoded by the reference nucleotide sequence. The term "substantially similar" is specifically intended to include nucleic acid molecules wherein the sequence has been modified to optimize expression in particular cells, e.g. in plant cells. The percentage of identity between the substantially similar nucleic acid molecule and the reference nucleotide sequence desirably is at least 45%, more desirably at least 65%, more desirably at least 75%, preferably at least 85%, more preferably at least 90%, still more preferably at least 95%, yet still more preferably at least 99%. Preferably, the percentage of identity exists over a region of the sequences that is at least about 50 residues in length, more preferably over a region of at least about 100 residues, and most preferably the sequences are substantially similar over at least about 150 residues. In a most preferred embodiment, the sequences are substantially similar over the entire length of the coding regions. Sequence comparisons may be carried out using a Smith-Waterman sequence alignment algorithm and as described in more detail below (see e.g. Waterman, M.S. Introduction to Computational Biology: Maps, sequences and genomes. Chapman & Hall. London: 1995. ISBN 0-412-99391-0, or at <http://www-hto.usc.edu/software/seqaln/index.html>). The local S program, version 1.16, is used with following parameters: match: 1, mismatch penalty: 0.33, open-gap penalty: 2, extended-gap penalty: 2.

Another indication that a nucleic acid sequences is a substantially similar nucleic acid of the invention is that it hybridizes to a nucleic acid molecule of the invention under stringent conditions. The phrase "hybridizing specifically to" refers to the binding, duplexing, or hybridizing of a molecule only to a particular nucleotide sequence under stringent conditions when that sequence is present in a complex mixture (e.g., total cellular) DNA or RNA. "Bind(s) substantially" refers to complementary hybridization between a probe nucleic acid and a target nucleic acid and embraces minor mismatches that can be accommodated by

reducing the stringency of the hybridization media to achieve the desired detection of the target nucleic acid sequence.

"Stringent hybridization conditions" and "stringent hybridization wash conditions" in the context of nucleic acid hybridization experiments such as Southern and Northern hybridizations are sequence dependent, and are different under different environmental parameters. Longer sequences hybridize specifically at higher temperatures. An extensive guide to the hybridization of nucleic acids is found in Tijssen (1993) *Laboratory Techniques in Biochemistry and Molecular Biology-Hybridization with Nucleic Acid Probes* part I chapter 2 "Overview of principles of hybridization and the strategy of nucleic acid probe assays" Elsevier, New York. Generally, highly stringent hybridization and wash conditions are selected to be about 5°C lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. Typically, under "stringent conditions" a probe will hybridize to its target subsequence, but to no other sequences.

The T_m is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly matched probe. Very stringent conditions are selected to be equal to the T_m for a particular probe. An example of stringent hybridization conditions for hybridization of complementary nucleic acids which have more than 100 complementary residues on a filter in a Southern or northern blot is 50% formamide with 1 mg of heparin at 42°C, with the hybridization being carried out overnight. An example of highly stringent wash conditions is 0.1 M NaCl at 72°C for about 15 minutes. An example of stringent wash conditions is a 0.2x SSC wash at 65°C for 15 minutes (see, Sambrook, *infra*, for a description of SSC buffer). Often, a high stringency wash is preceded by a low stringency wash to remove background probe signal. An example medium stringency wash for a duplex of, e.g., more than 100 nucleotides, is 1x SSC at 45°C for 15 minutes. An example low stringency wash for a duplex of, e.g., more than 100 nucleotides, is 4-6x SSC at 40°C for 15 minutes. For short probes (e.g., about 10 to 50 nucleotides), stringent conditions typically involve salt concentrations of less than about 1.0M Na ion, typically about 0.01 to 1.0 M Na ion concentration (or other salts) at pH 7.0 to 8.3, and the temperature is typically at least about 30°C. Stringent conditions can also be achieved with the addition of destabilizing agents such as formamide. In general, a signal to noise ratio of 2x (or higher) than that observed for an unrelated probe in the particular hybridization assay indicates detection of a specific hybridization. Nucleic acids that do not hybridize to each other under stringent conditions are

still substantially similar if the proteins that they encode are substantially similar. This occurs, e.g., when a copy of a nucleic acid is created using the maximum codon degeneracy permitted by the genetic code.

The following are examples of sets of hybridization/wash conditions that may be used to identify homologous nucleotide sequences that are substantially similar to reference nucleotide sequences of the present invention: a test sequence that hybridizes to the reference nucleotide sequence in 7% sodium dodecyl sulfate (SDS), 0.5 M NaPO₄, 1 mM EDTA at 50°C with washing in 2X SSC, 0.1% SDS at 50°C, more desirably in 7% sodium dodecyl sulfate (SDS), 0.5 M NaPO₄, 1 mM EDTA at 50°C with washing in 1X SSC, 0.1% SDS at 50°C, more desirably still in 7% sodium dodecyl sulfate (SDS), 0.5 M NaPO₄, 1 mM EDTA at 50°C with washing in 0.5X SSC, 0.1% SDS at 50°C, preferably in 7% sodium dodecyl sulfate (SDS), 0.5 M NaPO₄, 1 mM EDTA at 50°C with washing in 0.1X SSC, 0.1% SDS at 50°C, more preferably in 7% sodium dodecyl sulfate (SDS), 0.5 M NaPO₄, 1 mM EDTA at 50°C with washing in 0.1X SSC, 0.1% SDS at 65°C. The polynucleotide of the invention that hybridizes under the above conditions preferably comprises at least 80 base pairs, more preferably at least 50 base pairs and particularly at least 21, and more particularly 18 base pairs. Preferred homologs of use in the invention include nucleic acid molecules that encode an amino acid sequence that is at least 45% identical to SEQ ID NO:2, 6 or 8 as measured, using the parameters described below, wherein the amino acid sequence encoded by the homolog has trichothecene resistance activity, e.g. 3-O-acetyltransferase activity.

The term "substantially similar", when used herein with respect to a protein, means a protein corresponding to a reference protein, wherein the protein has substantially the same structure and function as the reference protein, e.g. where only changes in amino acids sequence not affecting the polypeptide function occur. When used for a protein or an amino acid sequence the percentage of identity between the substantially similar and the reference protein or amino acid sequence desirably is at least 45% identity, more desirably at least 65%, more desirably at least 75%, preferably at least 85%, more preferably at least 90%, still more preferably at least 95%, yet still more preferably at least 99%, using default BLAST analysis parameters and as described in more detail below.

Preferred homologs of the polypeptide of use in the invention comprise those having amino acid sequences that are at least 45% identical to SEQ ID NO:2, 6 or 8, wherein the

amino acid sequence encoded by the homolog has trichothecene resistance activity, e.g. 3-O-acetyl transferase activity.

Optimal alignment of nucleic acid or protein sequences for comparison can be conducted as described above and, e.g., by the local homology algorithm of Smith & Waterman, *Adv. Appl. Math.* 2: 482 (1981), by the homology alignment algorithm of Needleman & Wunsch, *J. Mol. Biol.* 48: 443 (1970), by the search for similarity method of Pearson & Lipman, *Proc. Nat'l. Acad. Sci. USA* 85: 2444 (1988), by computerized implementations of these algorithms (GAP, BESTFIT, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group, 575 Science Dr., Madison, WI), or by visual inspection (*see generally*, Ausubel *et al.*, *infra*).

One example of an algorithm that is suitable for determining percent sequence identity and sequence similarity is the BLAST algorithm, which is described in Altschul *et al.*, *J. Mol. Biol.* 215: 403-410 (1990). Software for performing BLAST analyses is publicly available through the National Center for Biotechnology Information (<http://www.ncbi.nlm.nih.gov/>). This algorithm involves first identifying high scoring sequence pairs (HSPs) by identifying short words of length W in the query sequence, which either match or satisfy some positive-valued threshold score T when aligned with a word of the same length in a database sequence. T is referred to as the neighborhood word score threshold (Altschul *et al.*, 1990). These initial neighborhood word hits act as seeds for initiating searches to find longer HSPs containing them. The word hits are then extended in both directions along each sequence for as far as the cumulative alignment score can be increased. Cumulative scores are calculated using, for nucleotide sequences, the parameters M (reward score for a pair of matching residues; always > 0) and N (penalty score for mismatching residues; always < 0). For amino acid sequences, a scoring matrix is used to calculate the cumulative score. Extension of the word hits in each direction are halted when the cumulative alignment score falls off by the quantity X from its maximum achieved value, the cumulative score goes to zero or below due to the accumulation of one or more negative-scoring residue alignments, or the end of either sequence is reached. The BLAST algorithm parameters W, T, and X determine the sensitivity and speed of the alignment. The BLASTN program (for nucleotide sequences) uses as defaults a wordlength (W) of 11, an expectation (E) of 10, a cutoff of 100, M=5, N=-4, and a comparison of both strands. For amino acid sequences, the BLASTP program uses as defaults

a wordlength (W) of 3, an expectation (E) of 10, and the BLOSUM62 scoring matrix (*see* Henikoff & Henikoff, *Proc. Natl. Acad. Sci. USA* 89: 10915 (1989)).

In addition to calculating percent sequence identity, the BLAST algorithm also performs a statistical analysis of the similarity between two sequences (*see, e.g.,* Karlin & Altschul, *Proc. Nat'l. Acad. Sci. USA* 90: 5873-5787 (1993)). One measure of similarity provided by the BLAST algorithm is the smallest sum probability (P(N)), which provides an indication of the probability by which a match between two nucleotide or amino acid sequences would occur by chance. For example, a test nucleic acid sequence is considered similar to a reference sequence if the smallest sum probability in a comparison of the test nucleic acid sequence to the reference nucleic acid sequence is less than about 0.1, more preferably less than about 0.01, and most preferably less than about 0.001.

A further indication that two nucleic acid sequences or proteins are substantially similar is that the protein encoded by the first nucleic acid is immunologically cross reactive with, or specifically binds to, the protein encoded by the second nucleic acid. Thus, a protein is typically substantially similar to a second protein, for example, where the two proteins differ only by conservative substitutions.

The phrase "specifically (or selectively) binds to an antibody," or "specifically (or selectively) immunoreactive with," when referring to a protein or peptide, refers to a binding reaction which is determinative of the presence of the protein in the presence of a heterogeneous population of proteins and other biologics. Thus, under designated immunoassay conditions, the specified antibodies bind to a particular protein and do not bind in a significant amount to other proteins present in the sample. Specific binding to an antibody under such conditions may require an antibody that is selected for its specificity for a particular protein. For example, antibodies raised to the protein with the amino acid sequence encoded by any of the nucleic acid sequences of the invention can be selected to obtain antibodies specifically immunoreactive with that protein and not with other proteins except for polymorphic variants. A variety of immunoassay formats may be used to select antibodies specifically immunoreactive with a particular protein. For example, solid-phase ELISA immunoassays, Western blots, or immunohistochemistry are routinely used to select monoclonal antibodies specifically immunoreactive with a protein. See Harlow and Lane (1988) *Antibodies, A Laboratory Manual*, Cold Spring Harbor Publications, New York ("Harlow and Lane"), for a description of immunoassay formats and conditions that can be

used to determine specific immunoreactivity. Typically a specific or selective reaction will be at least twice background signal or noise and more typically more than 10 to 100 times background.

"Conservatively modified variations" of a particular nucleic acid sequence refers to those nucleic acid sequences that encode identical or essentially identical amino acid sequences, or where the nucleic acid sequence does not encode an amino acid sequence, to essentially identical sequences. Because of the degeneracy of the genetic code, a large number of functionally identical nucleic acids encode any given polypeptide. For instance the codons CGT, CGC, CGA, CGG, AGA, and AGG all encode the amino acid arginine. Thus, at every position where an arginine is specified by a codon, the codon can be altered to any of the corresponding codons described without altering the encoded protein. Such nucleic acid variations are "silent variations" which are one species of "conservatively modified variations." Every nucleic acid sequence described herein which encodes a protein also describes every possible silent variation, except where otherwise noted. One of skill will recognize that each codon in a nucleic acid (except ATG, which is ordinarily the only codon for methionine) can be modified to yield a functionally identical molecule by standard techniques. Accordingly, each "silent variation" of a nucleic acid which encodes a protein is implicit in each described sequence.

Furthermore, one of skill will recognize that individual substitutions deletions or additions that alter, add or delete a single amino acid or a small percentage of amino acids (typically less than 5%, more typically less than 1%) in an encoded sequence are "conservatively modified variations," where the alterations result in the substitution of an amino acid with a chemically similar amino acid. Conservative substitution tables providing functionally similar amino acids are well known in the art. The following five groups each contain amino acids that are conservative substitutions for one another: Aliphatic: Glycine (G), Alanine (A), Valine (V), Leucine (L), Isoleucine (I); Aromatic: Phenylalanine (F), Tyrosine (Y), Tryptophan (W); Sulfur-containing: Methionine (M), Cysteine (C); Basic: Arginine (R), Lysine (K), Histidine (H); Acidic: Aspartic acid (D), Glutamic acid (E), Asparagine (N), Glutamine (Q). *See also*, Creighton (1984) *Proteins*, W.H. Freeman and Company. In addition, individual substitutions, deletions or additions which alter, add or delete a single amino acid or a small percentage of amino acids in an encoded sequence are also "conservatively modified variations."

A "subsequence" refers to a sequence of nucleic acids or amino acids that comprise a part of a longer sequence of nucleic acids or amino acids (e.g., protein) respectively.

Nucleic acids are "elongated" when additional nucleotides (or other analogous molecules) are incorporated into the nucleic acid. Most commonly, this is performed with a polymerase (e.g., a DNA polymerase), e.g., a polymerase which adds sequences at the 3' terminus of the nucleic acid.

Two nucleic acids are "recombined" when sequences from each of the two nucleic acids are combined in a progeny nucleic acid. Two sequences are "directly" recombined when both of the nucleic acids are substrates for recombination. Two sequences are "indirectly recombined" when the sequences are recombined using an intermediate such as a cross-over oligonucleotide. For indirect recombination, no more than one of the sequences is an actual substrate for recombination, and in some cases, neither sequence is a substrate for recombination.

A "specific binding affinity" between two molecules, for example, a ligand and a receptor, means a preferential binding of one molecule for another in a mixture of molecules. The binding of the molecules can be considered specific if the binding affinity is about $1 \times 10^4 \text{ M}^{-1}$ to about $1 \times 10^6 \text{ M}^{-1}$ or greater.

Substrate: a substrate is the molecule that an enzyme naturally recognizes and converts to a product in the biochemical pathway in which the enzyme naturally carries out its function, or is a modified version of the molecule, which is also recognized by the enzyme and is converted by the enzyme to a product in an enzymatic reaction similar to the naturally-occurring reaction.

Transformation: a process for introducing heterologous DNA into a cell, tissue, or insect. Transformed cells, tissues, or insects are understood to encompass not only the end product of a transformation process, but also transgenic progeny thereof.

"Transformed," "transgenic," and "recombinant" refer to a host organism such as a bacterium or a plant into which a heterologous nucleic acid molecule has been introduced. The nucleic acid molecule can be stably integrated into the genome of the host or the nucleic acid molecule can also be present as an extrachromosomal molecule. Such an extrachromosomal molecule can be auto-replicating. Transformed cells, tissues, or plants are understood to encompass not only the end product of a transformation process, but also transgenic progeny thereof. A "non-transformed," "non-transgenic," or "non-recombinant"

host refers to a wild-type organism, e.g., a bacterium or plant, which does not contain the heterologous nucleic acid molecule.

D) Summary

It is an object of the invention to provide a plant cell or cells comprising a heterologous polynucleotide encoding a gene product that is expressed in the plant cell wherein the gene product comprises trichothecene resistance activity.

Another object of the invention is to provide a plant comprising the above described plant cell wherein the plant is resistant to a trichothecene.

Another object of the invention is to provide a plant that is resistant to a trichothecene where the trichothecene comprises a C-3 hydroxyl group.

Another object of the invention is to provide a plant wherein the gene product is a 3-O-acetyltransferase.

Another object of the invention is to provide a plant of the invention wherein the heterologous polynucleotide is substantially similar to the nucleic acid sequence of SEQ ID NOs:1, 5 or 7.

Another object of the invention is to provide a plant of the invention wherein the heterologous polynucleotide comprises the nucleic acid sequence of SEQ ID NO:1, 5 or 7 or homologs thereof.

Another object of the invention is to provide a plant wherein the gene product is a polypeptide comprising a sequence substantially similar to SEQ ID NO:2, 6 or 8.

Another object of the invention is to provide a plant wherein the heterologous polynucleotide comprises the nucleic acid sequence of SEQ ID NO:1, 5 or 7.

Another object of the invention is to provide a plant comprising a heterologous polynucleotide, which comprises a consecutive 18 base pair portion identical in sequence to a consecutive 18 base pair portion set forth in SEQ ID NO:1, 5 or 7.

Another object of the invention is to provide a plant resistant to a trichothecene selected from the group consisting T-2 toxin, HT-2 toxin, isotrichodermol, 4,15-diacetoxyscirpenol (hereinafter "DAS"), 3-deacetylcalonectrin, 3,15-dideacetylcalonectrin, scirpentriol, neosolaniol; type B: 15-acetyldeoxynivalenol, nivalenol, 4-acetylnivalenol (fusarenone-X), 4,15-diacetylnivalenol, 4,7,15-acetylnivalenol, and DON.

Another object of the invention is to provide a plant resistant to DAS or DON.

Another object of the invention is to provide a seed of any of the plants of the invention.

Another object of the invention is to provide anyone of the above-described plants wherein the plant is a wheat, maize, barley or rice plant.

Another object of the invention is to provide a plant that is resistant to a fungus that produces a trichothecene comprising a C-3 hydroxyl group.

Another object of the invention is to provide a plant that is resistant to *Fusarium*, *Trichothecium* or *Myrothecium*.

Another object of the invention is to provide a plant that is resistant to *Fusarium*, in particular but not limited to *Fusarium graminearum*, *Fusarium culmorum*, *Fusarium sporotrichioides*, *Fusarium poae*, *Fusarium sambucinum*, *Fusarium equiseti*, *Fusarium acuminatum*, *Fusarium lateritium*, and *Fusarium pseudograminearum*.

Another object of the invention is to provide a plant that is resistant to *Fusarium graminearum*.

Another object of the invention is to provide a plant of the invention as described above wherein the heterologous polynucleotide is a microbial polynucleotide.

Another object of the invention is to provide a plant of the invention as described above wherein the microbial polynucleotide is a yeast or fungal polynucleotide.

Another object of the invention is to provide a plant of the invention as described above wherein the fungal polynucleotide is a *Fusarium* polynucleotide.

Another object of the invention is to provide a plant of the invention as described above wherein the *Fusarium* polynucleotide is a *Fusarium graminearum* or *Fusarium sporotrichioides* polynucleotide.

Another object of the invention is to provide a plant as described above wherein the plant is resistant to a fungus that produces a trichothecene.

Another object of the invention is to provide a plant as described above wherein the plant is resistant to a fungus that produces a trichothecene comprising a C-3 hydroxyl group.

Another object of the invention is to provide a method for producing a trichothecene resistant plant comprising the steps of:

- a) transforming a plant cell with a heterologous gene encoding a gene product, wherein the gene product increases resistance to a trichothecene; and
- b) expressing the gene product at a biologically significant level.
- c) regenerating the plant cell into a plant; and

- d) selecting a plant having increased resistance to a trichothecene.

Another object of the invention is to provide a method as described above further comprising the step of selecting a plant on which there is reduced growth of a fungus where the fungus produces a trichothecene.

Another object of the invention is to provide a method as described above wherein the fungus is of the genera *Fusarium*.

Another object of the invention is to provide a trichothecene resistant plant obtained according to the above-described methods.

Another object of the invention is to provide a seed produced by selfing or outcrossing a plant of the invention as described above, wherein a plant grown from the seed has an increased resistance to trichothecene.

Another object of the invention is to provide a method of preventing mycotoxin crop contamination comprising growing a plant of the invention as described above, wherein the plant is a crop plant.

Another object of the invention is to provide a method of preventing fungal growth on a crop, comprising growing a plant of the invention as described above, wherein the plant is a crop plant.

Another object of the invention is to provide a method of selecting transformed host cells, the method comprising: transforming a host cell with a nucleic acid construct encoding a trichothecene 3-*O*-acetyltransferase, and growing the transformed host cell in the presence of a trichothecene selective agent.

Another object of the invention is to provide a method of selecting transformed host cells wherein the host cells are plant cells, or microbial cells, particularly where the microbial cells are fungal cells.

Another object of the invention is to provide a method of selecting transformed host cells as described above where the host cell is further transformed with a second polynucleotide of interest.

Another object of the invention is to provide a method of selecting transformed host cells wherein the trichothecene is selected from the group consisting of T-2 toxin, HT-2 toxin, isotrichodermol, DAS, 3-deacetylcalonecetrin, 3,15-dideacetylcalonecetrin, scirpentriol, neosolaniol; type B: 15-acetyldeoxynivalenol, nivalenol, 4-acetylnivalenol (fusarenone-X), 4,15-diacetylnivalenol, 4,7,15-acetylnivalenol, and DON.

E) Detailed Description

Description of the Sequences:

- SEQ ID NO:1 is a cDNA sequence from *Fusarium sporotrichioides* encoding a polypeptide of the invention having trichothecene resistance activity.
- SEQ ID NO:2 is the polypeptide having trichothecene resistance activity encoded by SEQ ID NO:1.
- SEQ ID NO: 3 is a DNA primer.
- SEQ ID NO 4: is a DNA primer.
- SEQ ID NO: 5 is a DNA sequence from *Fusarium graminearum* encoding a polypeptide of the invention having trichothecene resistance activity.
- SEQ ID NO: 6 is the polypeptide having trichothecene resistance activity encoded by SEQ ID NO:5.
- SEQ ID NO. 7 is a DNA sequence from *Saccharomyces cerevisiae* encoding a polypeptide of the invention having trichothecene resistance activity.
- SEQ ID NO. 8 is the polypeptide having trichothecene resistance activity encoded by SEQ ID NO:7.
- SEQ ID NO. 9 is the DNA sequence of pCIB9818.
- SEQ ID NO. 10 is the DNA sequence of pAgroTRIr.
- SEQ ID NO. 11 is the DNA sequence of pNOV1704.

Description of the Drawing:

FIG. 1 depicts positions C-3 and C-8 on the representative trichothecene Deoxynivalenol.

The present invention relates to transgenic hosts particularly, transgenic plants, plant tissues, plant seeds, and plant cells comprising a heterologous polynucleotide encoding a gene product where the gene product comprises trichothecene resistance activity and methods of making and using the same. Trichothecene resistance activity as used herein refers to an activity that reduces or inhibits the phytotoxicity of a trichothecene, particularly to a fungus and/or plant, in a particular embodiment of the invention trichothecene resistance activity refers to an activity that transfers an acetate to the C-3 position (see Fig. 1) of a trichothecene.

The present invention further relates to transgenic hosts, particularly, transgenic plants, plant tissues, plant seeds, and plant cells expressing a heterologous polynucleotide encoding a gene product, the gene product having trichothecene resistance activity, particularly an acetyl transferase gene product, more particularly a 3-*O*-acetyl transferase gene product, more particularly trichothecene 3-*O*-acetyl transferase gene product and methods of making and using the same. Expression of the heterologous polynucleotide of the invention comprises the synthesis of RNA and may be detected by northern blot analysis. Particularly, expression of the heterologous polynucleotide of the invention may be detected where a labeled probe derived from a heterologous nucleotide of the invention, in particular embodiments, from SEQ ID NOs. 1, 5 or 7, hybridizes with RNA isolated from a transgenic plant of the invention in 7% sodium dodecyl sulfate (SDS), 0.5 M Sodium phosphate pH 7.0, 1 mM EDTA, 10 mg/ml BSA at 65°C with washing in .5% BSA (fraction V), 5% SDS, 40 mM Sodium phosphate pH 7.0, 1 mM EDTA, .25 M sodium chloride at 65 ° C, preferably in 1% SDS, 40 mM Sodium phosphate pH 7.0, 1 mM EDTA, .125 M sodium chloride at 65 ° C, and preferably in 1% SDS, 40 mM Sodium phosphate pH 7.0, 1 mM EDTA at 65 ° C.

The present invention further relates to transgenic plants plant tissues, plant seeds, and plant cells, expressing a heterologous polynucleotide of the invention where the plant, plant cell, plant tissue or plant seed is trichothecene resistant. Trichothecene resistant plants, plant cells, plant tissues and plant seeds as used herein are those which are capable of metabolism in the presence of a trichothecene which may be determined as described in Example 7 below. In a particular embodiment, trichothecene resistant plants, plant tissues, plant cells and plant seeds which have a specific enzyme activity of at least 10 nmol triacetoxyscirpenol (hereinafter "TAS")/microgram protein/15 min incubation at saturating substrate levels, more particularly at least 5 nmol TAS/microgram protein/15 min, more particularly at least 1 nmol TAS/microgram protein/15 min, more particularly at least .8 nmol TAS/microgram protein/15 min more particularly at least .5 nmol TAS/microgram protein/15 min, more particularly a specific activity of 0.25 nmol TAS/microgram protein/ 15 minute, more particularly a specific activity of .1 nmol TAS/microgram protein/15 min., more particularly a specific activity of .05 nmol TAS/microgram protein/15 min and even more particularly a specific activity of .01 nmol TAS/microgram protein/15 min above background levels of activity that occur naturally in a wild type control, particularly as determined in an assay as described in Example 6 below.

Trichothecene resistant plants of the invention comprise those of which a greater percentage of the seed germinate and form roots in the presence of a trichothecene than the seed from a wild type control where the trichothecene is present at a concentration of at least 5 microgram/ml, more preferably at least 10 microgram/ml, more at least preferably 15 microgram/ml, more preferably at least 20 microgram/ml and more preferably at least 25 microgram/ml. In a particularly preferred embodiment, trichothecene resistant plants of the invention comprise those of which at least 10% more seed, more preferably at least 20%, more preferably at least 30%, more preferably at least 40%, more preferably at least 50%, more preferably at least 60% more seed, more preferably at least 70% more seed, more preferably at least 80% more seed and more preferably at least 90% more seed germinate and form roots in the presence of a trichothecene than the seed of a wild type control.

Trichothecenes are frequently divided into several different structural groups. A particular embodiment of the present invention is drawn to resistance to group A and B trichothecenes. Groups A and B comprise the *Fusarium* trichothecenes and are differentiated primarily by the absence (group A) or presence (group B) of a carbonyl functional group at position C-8. FIG. 1 depicts the group B trichothecene, DON that, accordingly, comprises a carbonyl group at the C-8 position.

The present invention is more particularly drawn to resistance to trichothecenes, which contain a C-3 hydroxyl. Figure 1 depicts position C-3 on the representative trichothecene DON. Such trichothecenes include T-2 toxin, HT-2 toxin, isotrichodermol, DAS, 3-deacetylcalonecetrin, 3,15-dideacetylcalonecetrin, scirpentriol, neosolaniol; 15-acetyldeoxynivalenol, nivalenol, 4-acetylnivalenol (fusarenone-X), 4,15-diacetylnivalenol, 4,7,15-acetylnivalenol, and DON and their various acetylated derivatives.

In a particular embodiment, the trichothecene resistant plant, cell, tissue or seed thereof is resistant to a trichothecene producing fungus, particularly a fungus of the genera *Fusarium*. Fungus resistance as used herein refers to no initiation of infection after fungal inoculation or reduced spread of the infection after fungal inoculation compared to a wild type control.

In a preferred embodiment, a fungal resistant transgenic plant of the present invention is a cereal plant and under fungal challenge comprises less infected kernels or seeds compared to a wild type control, preferably at least a 10% decrease of infected kernels or seeds compared to the same number of kernels or seeds evaluated in a wild type control, more

preferably at least a 20% decrease, more preferably at least a 40% decrease and more preferably at least a 50% decrease of infected kernels compared to the same number of kernels or seeds in a wild type control. The fungal resistant transgenic cereal plants of the invention comprise but are not limited to maize, wheat, barley, rice, and oats.

In wheat, fungal spread in the head may be evaluated as described in Example 9 below, by counting the number of symptomatic and asymptomatic spikelets on each inoculated head and calculating the percentage of spikelets on each head that are symptomatic. In a preferred embodiment, fungal resistant wheat of the present invention comprises, under fungal challenge, less infected spikelets than the wild type control, preferably at least a 10% decrease of infected spikelets compared to the same number of spikelets evaluated in a wild type control, more preferably at least a 20% decrease, more preferably at least a 40% decrease and more preferably at least a 50% decrease of infected spikelets compared to the same number of spikelets in a wild type control.

In maize, fungal spread in the ear may be evaluated by visual estimation of the percentage of infected kernels as described further in Example 9 below. In a preferred embodiment, fungal resistant maize of the invention, under fungal challenge, comprise less infected kernels than the wild type control, preferably at least a 10% decrease in infected kernels compared to the number of infected kernels in the same number of ears visibly estimated in a wild type control, more preferably at least a 20% decrease, more preferably at least 30% decrease, more preferably at least a 40 % decrease and more preferably at least a 50% decrease in infected kernels compared to the same number of ears visibly estimated in a wild type control. In maize, internal fungal spread in the stalk may be visually evaluated by splitting open the stalk and assessing the amount of discoloration. In a preferred embodiment of the invention, the transgenic maize of the invention comprises less internal and/or external discoloration of the stalk compared to a wild type control.

In another, preferred embodiment fungal resistant plants of the invention comprise those of which a greater percentage of seed germinate in the presence of fungal challenge than germinate in the wild type control. In a particularly preferred embodiment, fungal resistant plants of the invention comprise those of which at least 10% more seed, more preferably at least 20%, more preferably at least 30%, more preferably at least 40%, more preferably at least 50%, more preferably at least 60% more seed, more preferably at least 70% more seed, more preferably at least 80% more seed and more preferably at least 90% more seed, more

preferably at least 100% more seed, more preferably at least 150% more seed germinates in the presence of *Fusarium* than does seed from the wild type control.

In another preferred embodiment, fungal resistant transgenic plants producing seed or kernels having less mycotoxin, e.g. trichothecene contamination, than the seed of a wild type control are provided. In a particularly preferred embodiment crop plants and more particularly cereal plants producing seed having at least 10% less trichothecene, more preferable at least 20% less trichothecene, more preferably at least 30% less trichothecene, more preferably at least 40% less trichothecene, more preferably at least 50% less trichothecene, more preferably at least 60% less trichothecene, more preferably at least 70% less trichothecene and more preferably at least 80% less trichothecene contamination than a wild type control are provided. Trichothecene contamination may be determined as described in Example 10 below.

The polynucleotides of use in the invention include heterologous polynucleotides encoding acetyl transferases, particularly those encoding acetyl transferases capable of conferring trichothecene resistance, more particularly those encoding trichothecene 3-O-acetyltransferases. In a particular embodiment, the heterologous polynucleotide of the invention may be derived from but is not limited to fungal origin, more particularly from *Fusarium*, *Trichothecium*, and *Myrothecium* origin, more particularly from a *Fusarium* species such as *F. acuminatum*, *F. crookwellense*, *F. culmorum*, *F. equiseti*, *F. graminearum* (*Gibberella zeae*), *F. lateritium*, *F. poae*, *F. sambucinum* (*G. pulicaris*), and *F. sporotrichioides*. Heterologous polynucleotides of use in the invention include SEQ ID NO:1, 5 and/or 7 and sequences substantially similar to SEQ ID NO:1, 5 and/or 7.

A polynucleotide of use in the invention can be incorporated into host cells, such as plant, fungal or bacterial cells, using conventional recombinant DNA technology. Generally, this involves inserting the polynucleotide into an expression system to which the polynucleotide is heterologous using standard cloning procedures known in the art. The vector contains the necessary elements for the transcription and translation of the polynucleotide of use in the invention in a host cell containing the vector. A large number of vector systems known in the art can be used, such as plasmids, bacteriophage viruses and other modified viruses. The components of the expression system may also be modified to increase expression. For example, truncated sequences, nucleotide substitutions, nucleotide optimization or other modifications may be employed. Expression systems known in the art can be used to transform virtually any crop plant cell under suitable conditions. A heterologous

polynucleotide of the inventions is preferably stably transformed and integrated into the genome of the host cells. In another preferred embodiment, the heterologous polynucleotide of the inventions is located on a self-replicating vector. Examples of self-replicating vectors are viruses, in particular Gemini viruses. Transformed cells can be regenerated into whole plants such that the chosen form of the polynucleotide of the invention confers trichothecene resistance in the transgenic plants.

I. Requirements for Construction of Plant Expression Cassettes

A polynucleotide of the invention intended for expression in transgenic plants is first assembled in an expression cassette behind a suitable promoter expressible in plants. The expression cassettes may also comprise any further sequences required or selected for the expression of the heterologous polynucleotide of the invention. Such sequences include, but are not restricted to, transcription terminators, extraneous sequences to enhance expression such as introns, vital sequences, and sequences intended for the targeting of the gene product to specific organelles and cell compartments. These expression cassettes can then be easily transferred to the plant transformation vectors described *infra*. The following is a description of various components of typical expression cassettes.

1. Promoters

The selection of the promoter used in expression cassettes will determine the spatial and temporal expression pattern of the heterologous polynucleotide of the invention in the transformed plant. Selected promoters will express heterologous polynucleotides of the invention in specific cell types (such as leaf epidermal cells, mesophyll cells, root cortex cells) or in specific tissues or organs (roots, leaves or flowers, for example) and the selection will reflect the desired location of accumulation of the gene product. Alternatively, the selected promoter may drive expression of the gene under various inducing conditions. Promoters vary in their strength, i.e., ability to promote transcription. Depending upon the host cell system utilized, any one of a number of suitable promoters known in the art can be used. For example, for constitutive expression, the CaMV 35S promoter, the rice actin promoter, or the ubiquitin promoter may be used. For regulatable expression, the chemically inducible PR-1 promoter from tobacco or *Arabidopsis* may be used (*see, e.g.*, U.S. Patent No. 5,689,044).

2. Transcriptional Terminators

A variety of transcriptional terminators are available for use in expression cassettes. These are responsible for the termination of transcription beyond the heterologous polynucleotide of the invention and its correct polyadenylation. Appropriate transcriptional terminators are those that are known to function in plants and include the CaMV 35S terminator, the *tm1* terminator, the nopaline synthase terminator and the pea *rbcS* E9 terminator. These can be used in both monocotyledonous and dicotyledonous plants.

3. Sequences for the Enhancement or Regulation of Expression

Numerous sequences have been found to enhance gene expression from within the transcriptional unit and these sequences can be used in conjunction with the polynucleotides of this invention to increase their expression in transgenic plants. For example, various intron sequences such as introns of the maize *Adhl* gene have been shown to enhance expression, particularly in monocotyledonous cells. In addition, a number of non-translated leader sequences derived from viruses are also known to enhance expression, and these are particularly effective in dicotyledonous cells.

4. Coding Sequence Optimization

The coding sequence of the selected gene optionally is genetically engineered by altering the coding sequence for optimal expression in the crop species of interest. Methods for modifying coding sequences to achieve optimal expression in a particular crop species are well known (see, e.g. Perlak *et al.*, *Proc. Natl. Acad. Sci. USA* 88: 3324 (1991); and Koziel *et al.*, *Bio/technol.* 11: 194 (1993); Fennoy and Bailey-Serres. *Nucl. Acids Res.* 21: 5294-5300 (1993). Methods for modifying coding sequences by taking into account codon usage in plant genes and in higher plants, green algae, and cyanobacteria are well known (see table 4 in: Murray *et al.* *Nucl. Acids Res.* 17: 477-498 (1989); Campbell and Gowri *Plant Physiol.* 92: 1-11(1990).

5. Targeting of the Gene Product within the Cell

Various mechanisms for targeting gene products are known to exist in plants and the sequences controlling the functioning of these mechanisms have been characterized in some detail. For example, the targeting of gene products to the chloroplast is controlled by a signal

sequence found at the amino terminal end of various proteins which is cleaved during chloroplast import to yield the mature protein (*e.g.* Comai *et al.* *J. Biol. Chem.* 263: 15104-15109 (1988)). Other gene products are localized to other organelles such as the mitochondrion and the peroxisome (*e.g.* Unger *et al.* *Plant Molec. Biol.* 13: 411-418 (1989)). The cDNAs encoding these products can also be manipulated to effect the targeting of heterologous products encoded by DNA sequences to these organelles. In addition, sequences have been characterized which cause the targeting of products encoded by DNA sequences to other cell compartments. Amino terminal sequences are responsible for targeting to the ER, the apoplast, and extracellular secretion from aleurone cells (Koehler & Ho, *Plant Cell* 2: 769-783 (1990)). Additionally, amino terminal sequences in conjunction with carboxy terminal sequences are responsible for vacuolar targeting of gene products (Shinshi *et al.* *Plant Molec. Biol.* 14: 357-368 (1990)). By the fusion of the appropriate targeting sequences described above to a heterologous polynucleotide of the invention, it is possible to direct a resulting product to any organelle or cell compartment.

B. Construction of Plant Transformation Vectors

Numerous transformation vectors available for plant transformation are known to those of ordinary skill in the plant transformation arts, and the polynucleotides pertinent to this invention can be used in conjunction with any such vectors. The selection of vector will depend upon the preferred transformation technique and the target species for transformation. For certain target species, different selection markers may be preferred. Selection markers used routinely in transformation include the *nptII* gene, which confers resistance to kanamycin and related antibiotics (Messing & Vierra, *Gene* 19: 259-268 (1982); Bevan *et al.*, *Nature* 304:184-187 (1983)), the *bar* gene, which confers resistance to the herbicide phosphinothricin (White *et al.*, *Nucl. Acids Res* 18: 1062 (1990), Spencer *et al.* *Theor. Appl. Genet* 79: 625-631 (1990)), the *hph* gene, which confers resistance to the antibiotic hygromycin (Blochinger & Diggelmann, *Mol Cell Biol* 4: 2929-2931), and the *dhfr* gene, which confers resistance to methotrexate (Bourouis *et al.*, *EMBO J.* 2(7): 1099-1104 (1983)), and the EPSPS gene, which confers resistance to glyphosate (U.S. Patent Nos. 4,940,935 and 5,188,642), phosphomannose isomerase gene, *manA*, which confers a selective metabolic advantage in the presence of mannose (U.S. Pat. Ser. No. 5,767,378 which is incorporated herein by reference in its entirety and Miles & Guest, *GENE*, 32:41-48 (1984)). PAT selectable marker that

confers resistance to BASTA (Sung H. Park et al., *In Vitro Cell.Dev.Biol.-Plant*, 34: 117-121 (1998)).

1. Vectors Suitable for *Agrobacterium* Transformation

Many vectors are available for transformation using *Agrobacterium tumefaciens*. These typically carry at least one T-DNA border sequence and include vectors such as pBIN19 (Bevan, *Nucl. Acids Res.* (1984)). Typical vectors suitable for *Agrobacterium* transformation include the binary vectors pCIB200 and pCIB2001, as well as the binary vector pCIB10 and hygromycin selection derivatives thereof. (See, for example, U.S. Patent No. 5,639,949).

2. Vectors Suitable for non-*Agrobacterium* Transformation

Transformation without the use of *Agrobacterium tumefaciens* circumvents the requirement for T-DNA sequences in the chosen transformation vector and consequently vectors lacking these sequences can be utilized in addition to vectors such as the ones described above which contain T-DNA sequences. Transformation techniques that do not rely on *Agrobacterium* include transformation via particle bombardment, protoplast uptake (e.g. PEG and electroporation) and microinjection. The choice of vector depends largely on the preferred selection for the species being transformed. Typical vectors suitable for non-*Agrobacterium* transformation include pCIB3064, pSOG19, and pSOG35. (See, for example, U.S. Patent No. 5,639,949).

C. Transformation Techniques

Once the polynucleotide of interest has been cloned into an expression system, it is transformed into a plant cell. Methods for transformation and regeneration of plants are well known in the art. For example, Ti plasmid vectors have been utilized for the delivery of foreign DNA, as well as direct DNA uptake, liposomes, electroporation, micro-injection, and microprojectiles. In addition, bacteria from the genus *Agrobacterium* can be utilized to transform plant cells.

Transformation techniques for dicotyledons are well known in the art and include *Agrobacterium*-based techniques and techniques that do not require *Agrobacterium*. Non-*Agrobacterium* techniques involve the uptake of exogenous genetic material directly by protoplasts or cells. This can be accomplished by PEG or electroporation mediated uptake,

particle bombardment-mediated delivery, or microinjection. In each case the transformed cells are regenerated to whole plants using standard techniques known in the art.

Transformation of most monocotyledon species has now also become routine. Preferred techniques include direct gene transfer into protoplasts using PEG or electroporation techniques, particle bombardment into callus tissue, as well as *Agrobacterium*-mediated transformation. Target tissue may be derived from such sources as wheat cultivar UC703 or maize genotype CG000526. For example, *Agrobacterium* mediated transformation of maize may be carried out as described in U.S. patent no. 6,162,965, which is herein incorporated by reference in its entirety which correspondingly published as WO 98/54961, and of barley may be carried out as described by: M. Cho, J. Wong, C. Marx, W. Jiang, P. Lemaux and B. Buchanan (1999). Overexpression of thioredoxin h leads to enhanced activity of starch debranching enzyme (pullulanase) in barley grain. PNAS 96: 14641-14646; S. Zhang, M. Cho, T. Koprek, R. Yun, P. Bregitzer and P. Lemaux (1999). Genetic transformation of commercial cultivars of oat (*Avena sativa* L.) and barley (*Hordeum vulgare* L.) using in vitro shoot meristematic cultures derived from germinated seedlings. Plant Cell Rep. 18: 959-966; P. Bregitzer, S. Harlbert and P. Lemaux (1998). Somaclonal variation in the progeny of transgenic barley. TAG 96: 421-425; M. Cho, W. Jiang and p. Lemaux (1998). Transformation of recalcitrant barley cultivars through improvement of regenerability and decreased albinism. Plant sci. 138: 229-244; P. Lemaux, m. Cho, S. Zhang, and p. Bregitzer (1998). Transgenic cereals: *Hordeum vulgare* L. – current status and future prospects. In: Vasil I, Phillips R (eds) Molecular Improvement of Cereal Crops, Kluwer Academic Publ, Dordrecht, The Netherlands, pp 255-316; S. Zhang, R. Williams-Carrier, D. Jackson, and P. Lemaux (1998). Expression of CDC2Zm and KNOTTED1 during in vitro axillary shoot meristem proliferation and adventitious shoot meristem formation in maize (*Zea mays* L.) and barley (*Hordeum vulgare* L.). Planta 204: 542-549; D. McElroy, J. Louwerse, S. McElroy and P. Lemaux (1997). Development of a simple transient assay for Ac/Ds activity in cells of intact barley tissue. Plant J. 11: 157-165; S. Tingay, D. McElroy, R. Kalla, S. Fieg, M. Wang, S. Thornton and R. Brettell (1997). *Agrobacterium tumefaciens*-mediated barley transformation. The Plant J. 11: 1369-1376; J. Qureshi, Z. Basri, R. Singh, R. Burton, M. Dalton, J. Kollmorgen and G. Fincher. 1988. *Agrobacterium*-mediated transformation of two varieties of barley (*Hordeum vulgare* L.) Proc. 42nd. Conference of Australian Society for Biochemistry and Molecular Biology, September 28-October 1, 1998, Adelaide, Australia; J.

Qureshi, R. Singh, Z. Basri, R. Stewart, R. Burton, J. Kollmorgen and G. Fincher (1997). Strategies for genetic transformation of elite Australian barley varieties. Proc. 8th. Aust. Barley Technical symp. Gold Coast, Queensland, 7-12 September 1997. 2:8.9-11; P. Lemaux, M. Cho, J. Louwerse, R. Williams and Y. Wan (1996). Bombardment-mediated transformation methods for barley. Bio-Rad US/EG Bull 2007: 1-6; T. Koprek, R. Hansch, A. Nerlich, R. Mendel and J. Schulze (1996). Fertile transgenic barley of different cultivars obtained by adjustment of bombardment conditions to tissue response. Plant Sci. 119: 79-91; T. Hagio, T. Hirabayashi, H. Machii and H. Tomutsune (1995). Production of fertile transgenic barley (*Hordeum vulgare* L.) plants using the hygromycin-resistance marker. Plant Cell Rep. 14: 329-334; H. Funatsuki, H. Kuroda, M. Kihara, P. Lazzeri, E. Muller, H. Lorz and I. Kishinami (1995). Fertile transgenic barley regenerated by direct DNA transfer to protoplasts. TAG 91: 707-712; A. Jahne, D. Becker, R. Bretschneider and H. Lorz (1994). Regeneration of transgenic, microspore-derived, fertile barley. TAG 89: 525-533; Y. Wan and P. Lemaux (1994). Generation of large numbers of independently transformed fertile barley plants. Plant Physiol. 104: 37-48.

II. Breeding

The polynucleotides of the invention can be utilized to confer trichothecene resistance to a wide variety of plant cells, including those of gymnosperms, monocots, and dicots. Although the heterologous polynucleotide of the invention can be inserted, e.g. transformed into any plant cell falling within these broad classes, it is particularly useful in crop plant cells, such as rice, wheat, barley, rye, corn, oats, potato, sweet potato, turnip, squash, pumpkin, zucchini, melon, soybean, and sorghum. The polynucleotides of use in the invention rendering a plant trichothecene resistant may be used in combination with other characteristics important for production and quality. The polynucleotides of the invention can be incorporated into plant lines through breeding approaches and techniques known in the art.

Where a trichothecene resistant gene allele is obtained by transformation into a crop plant or plant cell culture from which a crop plant can be regenerated, it is moved into commercial varieties using traditional breeding techniques to develop a trichothecene resistant crop without the need for genetically engineering the allele and transforming it into the plant.

III. Selection System

In another embodiment, the heterologous polynucleotide of use in the invention, can also be used as a selectable marker in transformation procedures. In this aspect the host cell is transformed with a second heterologous polynucleotide of interest as well as a heterologous polynucleotide of the invention which encodes a gene product comprising trichothecene resistance activity, using expressions cassettes and transformation techniques exemplified above and known in the art. After transformation, the transformed cells are selected for their ability to survive when exposed to a trichothecene, particularly DAS or DON or T-2 toxin. The host cell may be a eukaryotic or prokaryotic host cell using transformation and expression systems known in the art. The host cell may be a plant cell, a fungal cell, a bacterial cell, a yeast cell, an animal cell, or an insect cell.

In a particularly preferred embodiment of the invention, a polynucleotide which encodes a gene product comprising trichothecene resistance activity is used as a selectable marker in plant cell transformation methods. For example, plants, plant tissue, plant seeds, or plant cells expressing at least a second heterologous DNA sequence of interest can also be transformed to express a sequence encoding a polypeptide comprising a sequence substantially similar to that of SEQ ID NO:2, 6 or 8. The transformed cells are transferred to medium containing a phytotoxic trichothecene, particularly DAS and/or DON and/or T-2 toxin, in an amount sufficient to inhibit the growth or survivability of plant cells not expressing the polypeptide substantially similar to that having the amino acid sequence of SEQ ID NO:2, 6 or 8, wherein only the transformed cells will grow or will be unstunted. Concentrations of trichothecenes useful for selection of plants expressing the polypeptide substantially similar to that having the amino acid sequence of SEQ ID NO:2, 6 or 8 range from 1 ug/ml to 90 ug/ml. The method is applicable to any plant cell capable of expressing a polynucleotide comprising a nucleotide sequence substantially similar to that of SEQ ID NO: 1, 5 or 7, and can be used with any heterologous DNA sequence of interest. Expression of the second heterologous DNA sequence and the heterologous polynucleotide of the invention can be driven by the same promoter functional in plant cells, or by separate promoters.

F) Examples

The following examples further describe the materials and methods used in carrying out the invention and the subsequent results. They are offered by way of illustration, and their recitation should not be considered as a limitation of the claimed invention.

Example 1: Composition of pNOV1700, pCIB9818, pAgroTRIr and pNov 1704

1. pNOV1700:

pNOV1700 was deposited under the terms of the Budapest Treaty on March 19, 1999, with the Agricultural Research Service, Patent Culture Collection (NRRL), Northern Regional Research Center, 1815 Northern University Street, Peoria, Illinois 61604, USA and assigned accession number NRRL B-30117.

Accordingly, pNOV1700 comprises SEQ ID NO. 1 operably linked to the *Z.mays* ubiquitin promoter, including a portion of the exon and intron, and to the nopaline synthase polyadenylation signal.

2. pCIB9818

Plasmid pCIB9818 is a 6111 base pair circular plasmid having a DNA sequence according to SEQ ID NO. 9. The *Z. mays* ubiquitin promoter, base 12 to 1993 of SEQ ID NO.9, including a portion of the exon, base 896 to 1011, and the intron, base 1047 to 1993, is operably linked to the phosphate mannose isomerase selectable marker, base pair 2090 to 3193, the inverted PEPC intron #9 from base 3248 to 3355 and the termination sequence of the CaMV 35S gene, base 3357 to 3434.

3. pAgroTRIr

Plasmid pAgroTRIr is a 13,737 base pair circular binary vector having a DNA sequence according to SEQ ID NO. 10. Accordingly, pAgroTRIr comprises a selectable marker operable linked to a promoter and termination sequence and the polynucleotide region of SEQ ID NO: 1 behind and in frame with the Arabidopsis thaliana UBI 3 promoter (S. Norris, S. Meyer, and J. Callus, Plant Molecular Biology 22:895-906, (1993)) and in front of and in frame with the nos polyadenylation signal.

4. pNov1704

Plasmid pNOV1704 is a 12949 base pair circular binary vector having a DNA sequence according to SEQ ID NO. 11. The *Z. mays* ubiquitin promoter, base 11 to 1992 of SEQ ID NO.11(including exon 1 895 to 1010 and intron 1 1046 to 1992), is operably linked to the phosphate mannose isomerase selectable marker sequence, base 2089 to 3192, and the nopaline synthase termination sequence, base 3557 to 3688. pNOV1704, further comprises the *Z. mays* ubiquitin promoter,base 9218 to 11218 (including exon 1 10,110 to 10224 and intron 1 10225 to 11218) operably linked to the trichothecene 3-*O*-acetyl transferase sequence of SEQ ID NO.1, at base 11,234 to 12,662 and the nos termination sequence 12667 to 12935.

Example 2: Wheat transformation , selection and regeneration

Transformation

Immature zygotic embryos (0.75-1.25mm) are dissected from surface sterilized wheat caryopses (10% Chlorox X 10 minutes) then plated scutellum up onto an MS based medium (Murashige and Skoog, (1962) *Physiol. Plant* 15:473-439) supplemented with 3% sucrose, 3mg/liter 2,4-D (dichlorophenoxyacetic acid), 150mg/l glutamine, 75mg/l asparagine and solidified with 0.7% phytagar (3MS3S medium). The embryos are incubated in the dark at 28°C for 5-10 days prior to bombardment. The optimal time for bombardment is 6-7 days post-plating. Four hours prior to bombardment, the embryos are placed on a plasmolysis medium (same medium described above but with 15% maltose added in place of the sucrose) and arranged in a 2.5cm diameter circle with scutellum facing up.

pNOV1700, described in Example 1 above, is digested with *PvuII* and *XmnI* and a 4117bp fragment comprising a polynucleotide region having a sequence according to SEQ ID NO:1 as well as the ubiquitin promoter and NOS polyadenylation signal is isolated. pCIB9818, also described in Example 1 above, is digested with *AscI* and the 4246 bp fragment comprising the UBI maize promoter, selectable marker and CaMV 35S termination sequence is isolated.

The isolated DNA fragments are precipitated onto 0.3 micro meter gold particles using the standard Sanford method. While continuously vortexing , 5 microgram of the isolated fragment DNA per construct , 50ul of 2.5 M CaCl_2 and 20ul of 0.1M spermidine are added to an eppendorf tube that contains 50ul of 50% glycerol and 3mg gold. The DNA/gold mixture is given two ethanol washes. After discarding the supernatant from the last wash, ethanol is

added to make a final volume of 70 μ l. This provides six shots per tube of gold/DNA. The target plates are shot twice so that gives a delivery of approximately 3microgram DNA (if 5microgram each of 2 constructs is used which is usually the case) and 1.0 mg gold per target plate. The rupture pressure used is 1100 psi. After bombardment, the target plates are returned to the dark overnight. After approximately 24 hours of plasmolysis, the embryos are removed to 3MS3S and returned to the dark for 3 weeks of callus initiation. No subculturing is done during this time.

Selection/Regeneration

The embryogenic tissue that develops during the 3-week initiation period is dissected away from non-embryogenic tissue and placed on a regeneration/selection medium. The basic regeneration medium is 3MS3S without the 2,4-D but with 1mg/l GA3 (Gibberellin A₃) NAA (1-Naphtheleneacetic Acid) and NAA added instead (called NG medium). 10g/l mannose and 5g/l sucrose is added (NG1M.5S). The tissue is subjected to this initial phase of regeneration and selection for 2 weeks. For most of the 2 week period the tissue is in the light room. Shoot and root development begins during this phase and after 2 weeks all tissue is taken to the next stage.

For the second phase of regeneration and selection with mannose selection, mannose is decreased to 5g/l and the sucrose increased to 20g/l (MS2S.5M). The tissue normally stays on these media for approximately 4 weeks time during which further shoot and root development occurs.

Vigorously growing plantlets with good color, and root and shoot development are removed from plates and placed in larger containers called GA7's. This is the final stage of selection and regeneration. The medium contains only 1/2MS salts and 15g/l mannose. The best indicator that a plant may be transformed is the observance of active root growth into the medium. Leaf tissue from actively growing plantlets is collected and PCR is done for either the gene of interest or selectable marker before transferring to the green house.

EXAMPLE 3: Arabidopsis transformation

The binary vector pAgroTRlr constructs described in Example 1 above is transformed into *Agrobacterium tumefaciens* strain GV3101 (Bechtold, N. *et al.*, CR Acad. Sci. Paris, Sciences de la vie, 316:1194-1199 (1993)) by electroporation (Dower, W.J., Mol. Biol. Rep

1:5 (1987) A 25 ml culture from single colonies of GV3101 agrobacterium containing pAgroTRlr plasmids in YEB + Rifampin 100 and Kanomycin 100 is incubated at 30 degrees overnight. Large cultures are started by inoculating 500 ml of the same media with 5 mls of the small culture and are incubated overnight at 30 degrees. The OD at 600 nm of cultures is determined and the cultures are then spun down at 5 K in the GSA rotor for 15 minutes. Cells are resuspended in "IM Modified infiltration media" to achieve a final O. D. at 600 nm of .08. 200 microliters of Silwet per liter of suspended cells is added. Three pots of bolting Arabadopsis var Columbia about 4 plants per pot, are inverted in about 500 ml of cell suspension. The flowers are shaken in the cell suspension to dislodge the air bubbles and the plants are incubated in the cell suspension for 15 minutes. A dome is placed on the tray to keep the plants humid overnight.

Plants are allowed to grow about 3-4 weeks after which the plants are not watered for up to 1 week. Seed pods are collected and dried in drying room for about a week and a half. The seeds are planted and allowed to grow for about 2 weeks. The plants are sprayed with the selection agent and then sprayed again 2 days later and 4 days later. After about three days surviving plants can be transplanted to new pots.

EXAMPLE 4: Maize biolistic transformation.

Type I embryogenic callus cultures (Green et al 1983, Somatic cell genetic systems in corn. A. Fazelahmad, K. Downey, J. Schultz, RW Voellmy, eds. Advances in Gene Technology: Molecular Genetics of Plants and Animals. Miami Winter Symposium Series, Vol. 20. Academic Press, NY.) are initiated from immature maize embryos, that are 1.5 – 2.0 mm in length, from greenhouse grown material. Embryos are aseptically excised from surface-sterilized ears approximately 14 days after pollination. The embryos are placed on D callus initiation media (Duncan et al,(1985) Planta 165:pp322-332) with 2% sucrose and 5mg/L chloramben. Embryos and embryogenic cultures are subsequently cultured in the dark. Embryogenic responses are excised from the explants after about 14 days. Responses are placed onto D callus maintenance media with 2% sucrose and 0.5mg/L 2,4-D. After about 6 weeks of weekly selective subculture to fresh maintenance media, high quality compact embryogenic cultures are established. Actively growing embryogenic callus pieces are selected as target tissue for gene delivery. The callus pieces are plated onto target plates

containing maintenance medium with 12% sucrose approximately 4 hours prior to gene delivery.

The callus pieces are arranged in circles, with radii of 8 and 10mm from the center of the target plate.

pNOV1700, described in Example 1 above, is digested with PvuII and XmnI and a 4117bp fragment comprising a polynucleotide region having a sequence according to SEQ ID NO:1 isolated as well as promoter and polyadenylation signal. pCIB9818, also described in Example 1 above, is digested with AscI and the 4246 bp fragment comprising the marker gene, promoter and termination signal is isolated. The isolated DNA fragments are precipitated onto gold microcarriers as described in the DuPont Biolistics manual. Two to three μ g for each plasmid construct is used in each 6 shot microcarrier preparation. Polynucleotides of the invention are delivered to the target tissue cells using the PDS-1000He Biolistics device. The settings on the Biolistics device are as follows: 8 mm between the rupture disk and the macrocarrier, 10 mm between the macrocarrier and the stopping screen and 7 cm between the stopping screen and the target. Each target plate is shot twice using 650psi rupture disks. A 200 X 200 stainless steel mesh (McMaster-Carr, New Brunswick, NJ) is placed between the stopping screen and the target tissue. Seven days after gene delivery, target tissue pieces are transferred from the high osmotic medium to selection medium.

The target tissue is placed onto maintenance medium containing no sucrose and 1% mannose. After 3 to 5 weeks, growing callus pieces are subcultured to the maintenance medium containing no sucrose and 1.5% mannose. Embryogenic callus growing on selection media is subcultured every 2 weeks for 6 to 10 weeks until enough callus is produced to generate 10-20 plants. Tissue surviving selection from an original target tissue piece is subcultured as a single colony and designated as an independent transformation event. Colonies are transferred to a modified MS medium (Murashige and Skoog, 1962(1962) A revised medium for rapid growth and bioassays with tobacco tissue cultures. *Physiol. Plant* 15: 473-497.) containing 2% sucrose and 1% mannose (MS2S+1M) with 0.25mg/L ancyimido and 0.5mg/L kinetin. After 2 weeks, regenerating colonies are then transferred to MS2S+1M without hormones. Regenerating shoots with or without roots from all colonies are transferred to Magenta boxes containing MS3S medium and small plants with roots are recovered and transferred to soil in the greenhouse.

EXAMPLE 5: Analyses of transgenic plant expression

Tissue from transformed plants is analyzed for the presence of a polynucleotide comprising the sequence of SEQ ID NO:1. DNA is extracted from transformed plant and PCR analyses are performed according to standard protocols. The primers used for amplification of the gene constructs are (5'-acgaatcattaccgaggag-3') (SEQ ID No. 3) and (5'-ctcacactctcaggettacc-3') (SEQ ID NO. 4). A 650 nt fragment within the sequence of SEQ ID NO:1 in wheat obtained according to Example 2 above is detected.

b. Northern analysis

Transformed plants are analyzed for the presence of RNA by northern blot hybridization. For northern blot analysis, RNA extracted from plant tissue is size separated and blotted onto a nylon membrane. This membrane is subsequently hybridized with a radioactive probe, derived from the 429 nt Styl fragment of the polynucleotide according to SEQ ID NO:1 is used as the probe. RNA is detected in wheat and arabadopsis plants transformed according to examples 2 and 3 above.

EXAMPLE 6: Enzymatic assay for trichothecene 3-O-acetyltransferase activity.

1. a.) **Extraction of plant tissue for enzyme assays:** Three 1 x 1/8 in pieces of leaf (about 50 mg) from transgenic plants of the invention including those transformed and regenerated according to Examples 2-4 above are selected.
(b) Glass Bead Mill: Tissue is placed in 2 ml round bottomed tube and the cap closed. The tube is immersed in liquid nitrogen and is incubated overnight at -80 °C. Tube is shaken on saws-all 24 seconds and 0.4 ml sodium phosphate buffer is added. The tube is vortexed about 10 seconds and is placed on ice. The tube is vortexed another 5 minutes and then is spun at 14,000 rpm in Eppendorf centrifuge 5 min. The supernatant is removed and is placed in a clean tube.
2. a) The following components are mixed
trichothecene substrate, 2 microliters of DAS (20% acetone in 50 mM Sodium phosphate buffer pH 7.0). DON may also be used.
Acetyl CoA substrate, 2 micro liters of [¹⁴C]-acetyl CoA NEN cat. # NEC313 (60 mCi/millimole and 0.02 mCi/ml)
Buffer, to a final volume of 50 µl with sodium phosphatate buffer pH 7.0

b) The assay is initiated by adding the following enzyme preparation and is incubated at 30 °C for 15 minutes.

Enzyme preparation, 10 microliter plant extract in sodium phosphate buffer pH 7.0

c) After 15 minutes, 100 microliters ethyl acetate is added and the tube is vortexed twice for several seconds. The tube is spun for 2 minutes at 14,000 rpm in an Eppendorf centrifuge. 50 microliters of the ethyl acetate phase is removed and is added to a vial containing scintillation cocktail. The tube is counted for 2 min. using a scintillation counter.

Twenty separate wheat plants obtained according to Example 2 above and having specific activities of 0.60 to 13.4 nmol acetylated product/ug protein/15 min are identified. The specific activities of the transformed wheat plants are significantly greater than the negative control. The negative control is a non-transformed wheat cultivar, which has a specific activity of 0.1 to 0.2 nmol acetylated product/ug protein/15 min.

Five separate *Arabidopsis* plants obtained according to Example 3 above and having specific activities ranging from 3.8 to 28 nmol acetylated product/ug protein/15 min are identified. The specific activities of the transformed plants are significantly greater than that of the negative control. The negative control is an *Arabidopsis thaliana* var columbia transformed with a nucleic acid construct for expressing the selectable marker which has a specific activity of less than 0.1 nmol acetylated product/ug protein/15 min.

Maize plants from at least two different transformants obtained according to Example 4 above and having specific activities ranging from 11.1 to 17.9 nmol acetylated product/ug protein/15 min are identified. The specific activities of the transformed plants are significantly greater than that of the negative control. The negative control is a non-transformed *maize* genotype that has a specific activity of less than 0.2 nmol acetylated product/ug protein/15 min.

Maize plants from at least 16 different transformants obtained using *Agrobacterium* mediated transformation of pNOV 1704, having specific activities ranging from 17 to 183 nmol/microgram/15 min are identified.

EXAMPLE 7: Bioassay for trichothecene resistance in transgenic plants.

250 ml of CPR media having the following components is prepared and the pH adjusted to 6.5 with KOH.

½ MS salts 0.54 g

½ MS vitamins 1.25 ml

Sucrose 1% (optional) 2.50 g

Agarose is added to the above media to a concentration of 1% (2.50 g) and the media is autoclaved. 25 ml of 50 mg/ml chlorophenol red is added to the autoclaved media.

While media is maintained at 55 °C, DAS or DON is added in acetone at various concentrations. (i. e. DON at 4, 8, or 16 microliters 10 mg/ml or DAS at 2, 4, or 6 microliters 50 mg/ml DAS per 1.7 ml). About 0.5 ml of media is aliquoted to each well in a 48 well microtiter plate.

1/3 x 1/8 inch pieces of transformed plant tissue are added to microtiter plate wells as well as control tissue from untransformed wild type controls. The leaf pieces are allowed to fall into a petri dish and are pushed into the microtiter plate well media with tweezers. The microtiter plates are incubated 2 to 4 days at 20 °C under lights. Leaf piece metabolism results in color change (drop in pH) from red to yellow. Trichothecene resistance activity or reduced sensitivity to trichothecenes by transformants, results in yellow colored wells in the presence of DAS or DON.

A color change from red to yellow compared to the control that remains red is observed in wheat and maize plants of the invention. Furthermore, the individual leaf pieces have significantly less chlorosis than the corresponding control.

EXAMPLE 8:Germination Assay

A. Trichothecene Resistance Germination Assay

Seed from transgenic plants of the invention is grown under selective pressure from the selection agent and the resulting plants are selfed. The resulting seed is plated on MS3S medium (MS salts 4.3 g/L, MS vitamins 100X, Sucrose 30 g/L, and phytagar 8 g/L) and supplemented with either DAS or DON (at 20 mg/ml) at a density of 1000 to 1200 seeds/petri dish (100 mm diameter). After incubation in the light for four days the plates are examined for seedling growth.

Arabidopsis seed from plants obtained according to Example 3 above and grown in media comprising DAS, has numerous plants with both root and shoot development. While control seed (parental Arabidopsis line, var. Columbia) germinates poorly and no roots form when grown in DAS supplemented media. No differences are observed between transformed and control seeds grown in the same media without DAS.

B. Fungal Resistance Germination Assay for Detecting Resistance to Seedling Blight

1. Wheat Fungal Resistance Germination Assay:

Fungal resistance germination assays in wheat are carried out substantially as described by R. H. Proctor, T. M. Hohn, and S. P. McCormick. Reduced virulence of *Gibberella zeae* caused by disruption of a trichothecene toxin biosynthetic gene. *Mol.Plant-Microbe Interact.* 8 (4):593-601, 1995.) which is herein incorporated by reference in its entirety.

Inoculum consists of macroconidia of *F. graminearum* diluted in water to 1×10^6 conidia per ml. Inoculum is prepared by washing the macroconidia from V-8 juice agar cultures grown under white and near UV fluorescent lights for 7-10 days. In seedling assays, seeds of two different transgenic wheat events from Example 2 above and the wild type control are surface sterilized by washing in a 10% bleach and .05% Tween solution for approximately 15 min. and rinsed five times with sterile distilled water. The seeds are soaked in a suspension of macroconidia for approximately 10 min and then sown in vermiculite contained in 10 cm plastic pots (20 seeds per pot). Prior to sowing, the pots are filled approximately $\frac{3}{4}$ full with vermiculite and set in 2-4 cm of water until the top of the vermiculite was wet. After sowing, seeds are covered with an additional 1-2 cm of vermiculite and pots are placed individually into plastic bags and incubated in a growth chamber at 22 °C with 16 h light and 8 h dark for week. After approximately one week the pots are removed from the bags, and after two weeks, disease is evaluated by counting the number of seedlings that emerge in each pot. Controls are treated as described above except that the seeds are soaked in sterile water and 40 seeds are used.

50% and 43% of the seed from the two different transgenic plant events germinate as compared to the same transgenic seed treated with water, whereas, 17% of the wild type control germinate compared to the same seed treated with water.

2. Maize Fungal Resistance Germination Assay:

Inoculum is produced from *F. graminearum* cultures grown on mung bean agar (made with liquor from boiled mung beans) under 12h alternating light and dark cycles at 25°C. Spores are harvested by first flooding the plate with sterile water and then scraping the plate using a glass rod. The solution is collected and the spore concentration adjusted to 1×10^6 spores/ml with double distilled, sterile water on the day of inoculation.

Soil consisting of a sterilized mix of soil, peat, and vermiculite is inoculated with 1 ml of spore solution/liter of soil in 5 liter flats and the inoculated soil is mixed in a cement mixer for 2 minutes per load. Control treatments consist of non-infested soil. Flats are planted with 30 kernels each of the transgenic seed of the invention or wild type control and incubated in growth chambers kept at 55°F under semi-saturated soil conditions for up to 4 weeks. Light levels are under 24-hour darkness for 14 days post planting, and 12-hour light 15-24 days post planting. Labeled stakes are added to flats, flats are moved to the growth chamber and randomized.

Plant counts are initiated as soon as emergence begins, and are performed daily or every other day until there is no longer a change in plant emergence.

Symptoms of visual discoloration and damping-off of seedling emergence are determined and used to characterize the degree of plant resistance as compared to wild type controls. Plants having less visual discoloration and/or damping off of seedling emergence than the wild type control are selected.

EXAMPLE 9: Fungal Resistance Assay

A. Testing of transgenic wheat plants

Wheat head blight or head scab is caused by *Fusarium graminearum* (teleomorph: *Gibberella zeae*). *F. graminearum* cultures are grown on V-8 agar medium (made with V-8 juice) or on mung bean agar (made with liquor from boiled mung beans) under 12h alternating light and dark cycles at 25°C. Spores are harvested by first flooding the plate with sterile water and then scraping the plate using a glass rod. The solution is collected and the spore concentration adjusted to 5×10^4 spores/ml with double distilled, sterile water on the day of inoculation.

Transgenic plants are obtained as described in Example 2 above, and control plants may be grown in the greenhouse until anthesis or heading. Heads are inoculated by injecting

approximately 20 ul (about 1000 spores) of inoculum between the lemma and palea of one floret near the middle of each head. Some heads are left uninoculated or are inoculated with sterile water as controls. Plants are then moved to a growth chamber and incubated under high humidity for up to 21 days or plants are first incubated in a mist chamber for 72 h at 65 to 70° F and then incubated in the greenhouse for an additional 18 days.

Transgenic plants of the invention and control plant may also be grown in the field, where heads are inoculated by spraying. Disease is evaluated by counting the number of symptomatic and asymptomatic spikelets on each inoculated head on a representative number of transgenic heads and wild type control heads and from this calculating the percentage of spikelets on each head that are symptomatic. Symptoms consist of premature whitening as compared to the control plants and in some cases necrosis of spikelets. Plants having fewer symptomatic spikelets than the wild type control are selected.

Six different transgenic plants having different copy numbers of SEQ ID NO.1 and different numbers of insertion sites according to southern data have average percent of symptomatic spikelets per head ranging from 10.40% to 31.20% compared to 44.75% for the wild type control, where the transgenic plants and controls are incubated in the greenhouse as described above. These same transgenic plants have enzymatic activities, measured as described in example 6 above, ranging from .874 to 29.1 nmol/microgram/15 min.

B. Testing of transgenic corn plants

Corn ear rot assay.

Corn ear rot is caused by *Fusarium graminearum* (teleomorph: *Gibberella zeae*). *F. graminearum* cultures are grown on V-8 agar medium (made with V-8 juice) under 12h alternating light and dark cycles at 25°C. Spores are harvested by first flooding the plate with sterile water and then scraping the plate using a glass rod. The solution is collected and the spore concentration adjusted to 5×10^5 spores/ml with double distilled, sterile water) on the day of inoculation. Transgenic plants and control plants are grown in the greenhouse or field. Where grown in the greenhouse, the transgenic and control plants are maintained in the greenhouse until four to seven days post silk emergence when a 2 ml spore suspension is introduced into the silk channel (inside the husk cavity and above the cob). This is accomplished using an 18-gauge stainless steel hypodermic needle attached to a large syringe. In addition to silk

channel inoculations, a kernel inoculation method is also used to assay disease resistance. Kernel inoculation involves the introduction of the spore suspension (approx. 0.4 ml) into a group of four kernels through multiple injections with an 18-gauge needle attached to a syringe. Disease is evaluated by visual inspection of ears harvested 5 to 7 weeks post-inoculation for visibly infected kernels. The disease rating scale for husked ears is based on a visual estimation of the percentage of visibly infected kernels on an ear as follows: 1 is 0%; 2 is 1 to 3%; 3 is 4 to 10%; 4 is 11 to 25%; 5 is 26 to 50%; 6 is 51 to 75%; 7 is 76 to 100%. Maize plants are selected that have a lower percentage of visibly infected kernels compared to the wild type control.

EXAMPLE 10: Mycotoxin Contamination Assay

Samples are prepared for mycotoxin concentration analysis as follows. Seed is collected from transgenic plants of the invention weighed and bulked together. Where wheat seed is being assayed, wheat seed is collected from the heads of the same transgenic plants of the invention and weighed and bulked together. Where transgenic maize is being assayed, corn ears are dried to low moisture levels, ears are hand-shelled and kernels from ears of the same transgenic plant are weighed and bulked together. Each seed or kernel sample is mixed thoroughly to promote a random distribution of seed. A 50 g seed or kernel sample is ground to a fine powder in a mill (e.g. Retsch ultra centrifugal mill type ZM1, Brinkman Instruments, Inc., Rexdale, Ontario, Romer Series II Mill, Union, Missouri, USA). The concentration of the mycotoxin of interest such as, DON is then determined using the commercially available tests such as DONtest TAG™ mycotoxin testing system (VICAM, LP, 313 Pleasant Street, Watertown, MA 02472) or analyzed by a commercial analysis company (e.g. Romer Labs, Inc, Union, Missouri, USA or Trilogy Analytical Laboratory, Inc., Beaufort, Missouri, USA). The manufacturer's instructions are followed for all aspects of the analysis. For DONtest TAG™ mycotoxin testing system, a final fluorometric measurement for DON is conducted. Plants producing seed or kernels having less mycotoxin, such as DON, than the wild type control are selected.

Example 11: Use of polynucleotide according to SEQ ID NO:1 as a selectable marker.

A. Selectable Marker in Fungal Cells.

Ashbya gossypii is transformed using standard fungal transformation techniques with a DNA construct comprising a polynucleotide having the sequence of SEQ ID NO:1 operably linked to the galactosidase promoter. Transformed cells grow in media comprising DAS at a concentration ranging from 1.56ng/ml to 196 pg/ml whereas as the untransformed wild type fungal cells do not.

B. Selectable Marker in Plant Cells.

Seed from *Arabidopsis* plants transformed according to Example 3 above but not yet subjected to selection is plated out in .1% agarose medium containing 0, 5, or 10 ug/ml DAS. After incubation in a growth room at 22 C with 16 hours of light and 8 hours of darkness for 2 weeks, the larger unstunted plants are transplanted from a DAS plate, and a corresponding number are transplanted from the control plate.

Leaves of *Arabidopsis* plants transplanted from the 5 microgram/ml plate, are assayed for enzymatic activity after a 2 week growth period, and showed 11 out of 11 unstunted plants were enzymatically active as measured by Example 6 while 9 out of 10 plants not selected by DAS were negative in the same assay. The one non-selected plant that was enzymatically active was much less active than any of the DAS selected plants assayed.

The above-disclosed embodiments are illustrative. This disclosure of the invention will place one skilled in the art in possession of many variations of the invention. All such obvious and foreseeable variations are intended to be encompassed by the appended claims.